

Impact Factor – 6.261 • Special Issue - 120 • Feb, 2019 • ISSN – 2348-7143

INTERNATIONAL RESEARCH FELLOWS ASSOCIATION'S

## **RESEARCH JOURNEY**

UGC Approved Journal  
Multidisciplinary International E-research Journal

*National Conference On*

# **RECENT DEVELOPMENTS IN LIFE SCIENCES FOR HUMAN WELFARE**

... Organized by ...

*Department of Botany*

**Yeshwantrao Chavan College of Arts, Commerce and Science, Sillod.**

Dist. Aurangabad, 431112 (M.S.) India

... Guest Editor ...

**Principal Dr. Ashok Pandit**

... Chief Editor ...

**Dr. Dhanraj T. Dhangar**

... Executive Editors ...

**Dr. Smt. Sunita Jawale**

**Dr. J. H. Sawdekar | Dr. K. V. Badar**

*Printed by :* PRASHANT PUBLICATIONS, JALGAON

Impact Factor – 6.261 ▪ Special Issue - 120 ▪ Feb. 2019 ▪ ISSN – 2348-7143

INTERNATIONAL RESEARCH FELLOWS ASSOCIATION'S

## **RESEARCH JOURNEY**

UGC Approved Journal

Multidisciplinary International E-research Journal

Printed by

### **PRASHANT PUBLICATIONS**

3, Pratap Nagar, Sant Dnyaneshwar Mandir Road, Near Nutan Maratha Mahavidyalaya, Jalgaon.

Website: [www.prashantpublications.com](http://www.prashantpublications.com) Email: [prashantpublication.jal@gmail.com](mailto:prashantpublication.jal@gmail.com)

Ph: 0257-2235520, 2232800, 9665626717, 9421636460

**EDITORIAL POLICIES** - Views expressed in the papers / articles and other matter published in this issue are those of the respective authors. The editor and associate editors does not accept any responsibility and do not necessarily agree with the views expressed in the articles. All copyrights are respected. Every effort is made to acknowledge source material relied upon or referred to, but the Editorial Board and Publishers does not accept any responsibility for any inadvertent omissions.

## Amylase Activity of Saprophytic Fungi from Vegetables Wastes.

Balwan W. Kamble, Shrimant A. Survase and Anil U. Kulkarni

Department of Botany, Lalbahadur Shastri Senior College,  
Partur Dist. Jalna.431501 (M.S.) India

### Abstract :

Fungal Amylase is used in commercial starch modification, cleaning compounds, textile processing, fermentation pretreatment, and animal feed supplements. In present study the production of amylase by ten dominating fungal species associated with vegetable waste were tested on substrate (starch nitrate) and non substrate (glucose nitrate) medium. Amylase activity was determined by cup plate method. Among ten dominating fungi selected for the study. Fungi show maximum amylase production on the basis of zone of clearance on the cup plate method. The results showed that maximum enzyme production obtained by *Aspergillus fumigatus*, *Fusarium oxysporium* and *Rhizopus stolonifer* as compare to other fungi like *Aspergillus niger*, *A. flavus*, *Alternaria alternata*, *Curvularia lunata*, *Penicillium notatum*, *P. spp.* and *Trichoderma viride*.

**Key word :** Substrate, non substrate medium, cup plate method, fungi, amylase activity, vegetable waste.

### Introduction :

Amylases are a group of important enzymes which are mainly employed in the starch processing industries for the hydrolysis of polysaccharides like starch into simple sugars (Akpan et al., 1999; Damien et al., 2010). Amylases are one of the most important industrial enzymes that have a wide variety of applications ranging from conversion of starch into sugar syrups, to the production of cyclodextrins for the pharmaceutical industry. Amylases accounts for about 30% of the world's enzyme production ((Vander et al., 2002; Rita et al., 2009). Amylases are among the most important enzymes which hydrolyze starch molecules to give diverse products including dextrin and progressively smaller polymers composed of glucose unit (Gupta et al., 2009).

The microorganisms have used for a huge importance to textile, food, baking and detergent industries and excite a large interest into the exploration of enzyme activity in microorganisms (Sivaramakrishnan et al., 2006). Amylases from plants and microbial sources have been employed for centuries in brewing industry (Pal and Khanum 2010). Fungal amylases are widely used for the preparation of oriental foods (Hernandez et al., 2006). Fungal amylases especially *Aspergillus niger* has been concentrated because of their ubiquitous nature and non fastidious nutritional requirement [Abu et al., 2005]. Ikenebomeh Chickundu in the year 1997, reported that *A.niger* shows high amyolytic activity in biomass production. Submerged fermentation holds tremendous potential for the production of enzymes.

The diversity of amylases, in contrast to the specificity of their action, attracted worldwide attention in attempts to exploit their physiological and

biotechnological applications (Wang et al., 2011) Starch degrading enzymes like amylase have received great deal of attention because of their perceived technological significance and economic benefit.

The present work is undertaken for screening of amylase producing fungi. From vegetable waste and optimization of cultural conditions.

### Materials and Methods :

Ten fungi were used that is *Aspergillus niger*, *A.flavus*, *A. fumigatus*, *Alternaria alternata*, *Curvularia lunata*, *Penicillium notatum*, *P. spp.*, *Rhizopus stolonifer* and *Trichoderma viride*, were used. These fungi were tested for *in-vitro* production of Amylase enzymes activity.

### Production of Amylase :

Detection of amylase was also studied in the present work. The isolated fungi were grown on liquid medium containing starch 1%, KNO<sub>3</sub> - 0.25 %, KH<sub>2</sub>PO<sub>4</sub> - 0.1% and MgSO<sub>4</sub>.7H<sub>2</sub>O - 0.05 %. Twenty five ml of the medium was poured in 100 ml conical flasks. The flask along with medium was sterilized by autoclaving at 15 lbs pressure for 20 minutes. The autoclaved flasks were allowed to cool and isolated fungi were inoculated with 1 ml of spore suspension which was prepared from freshly growing cultures on PDA slants. The inoculated flasks were incubated at 27 ± 2 °C for 7 days. After 7 day the flasks were harvested and the contents were filtered through Whatman filter paper No. 1. The filtrate were collected in pre sterilized conical flasks and termed as crude enzyme preparation.

### Amylase assay for (Cup-plate method) :

The amylase activity was determined with the help of cup late method as described by Danai (1994). In this method, 20 ml of starch nitrate medium (substrate) starch

1% and agar 2% were used. The starch assay medium after sterilization was poured in each pre sterilized petriplates. The medium is allowed to cool and solid and after this a cavity was made with the help of a cork borer (8mm). The cavity was filled with 1 ml culture filtrate (crude enzyme). The plates were incubated at 27 °C for 24 hours. The plates were flooded with Lugols iodine solution as an indicator and kept it for 20 - 40 minutes. A distinct activity zones are clearly seen after removing the iodine solution with distilled water. The clear zone was resulted due to the activity of amylase and the diameter of this zone was measured and noted. Similar procedure followed for the glucose nitrate medium (non substrate).

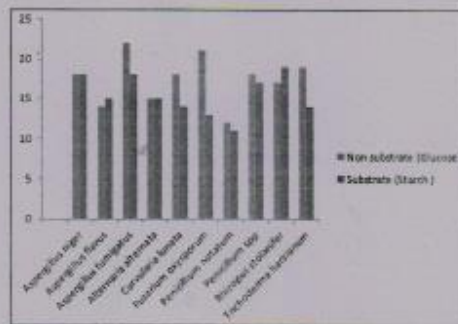
**Results and Discussion :**



**Table : Production of amylase on substrate and non substrate media**

Fungi	Non substrate (Glucose)	Substrate (Starch)
Zone diameter in mm		
<i>Aspergillus niger</i>	18	18
<i>Aspergillus flavus</i>	14	15
<i>Aspergillus fumigatus</i>	22	18
<i>Alternaria alternata</i>	15	15
<i>Curvularia lunata</i>	17	14
<i>Fusarium oxysporum</i>	21	13
<i>Penicillium notatum</i>	12	11
<i>Penicillium spp.</i>	18	17
<i>Rhizopus stolonifer</i>	17	19
<i>Trichoderma harzianum</i>	19	14

**Graph : Production of amylase by saprophytic fungi.**



Detection of extracellular amylase was carried out in the same experiment and all the tested fungi secreted amylase in the medium. From the result highest amylase activity was detected in non- substrate broth medium from by *Aspergillus fumigatus* (22mm) followed by *Fusarium oxysporum* (21mm). Moderate amylase activity was detected in *Trichoderma viride*, *Aspergillus niger* *Penicillium species*, *Rhizopus stolonifer*, *Alternaria alternata*, *Aspergillus flavus* and *Penicillium notatum*. In substrate medium maximum amylase activity exhibited by *Rhizopus stolonifer* (19mm) followed by *Aspergillus niger* (18mm) and *A. fumigatus* (18mm). However, *Penicillium species*, *Aspergillus flavus*, *Alternaria alternata*, *Trichoderma viride* *Curvularia lunata*, *Fusarium oxysporum* and *Penicillium notatum* had a least significant amylase activity. Same result found by Kulkarni et.al, (2012) and Sulochana et. al, (2010).

It can concluded from table and graph, that amylase enzyme production was found to be maximum in both substrate and non substrate broth media for all tested fungi in general. *Aspergillus fumigatus* and *Fusarium oxysporum* showed highest amylase production on non substrate medium compared to the other fungi. Substrate medium also respond to high amylase production by *Rhizopus stolonifer*. Minimum production of amylase was seen in *Penicillium notatum*. From Vegetables waste, saprophytic fungi were highly beneficial for the production of amylase.

**Acknowledgement :**

Authors are thankful to Principal of Lalbahadur Shastri Sr Colleges Partur for providing all necessary facilities.

**Reference :**

1. Akpan I. Bankole Mo, Adesemowo Am (1999). A rapid plate culture method for screening of amylase producing microorganisms. *Biotechnol. Tech.*13:411-413.
2. A. Pal and E. Khanum (2010). "Production and extraction optimization of xylanase from *Aspergillus niger* DFR-5 through solid-state-fermentation," *Journal of Bioresource Technology*, vol 101, no. 19, pp. 7563-7569.
3. Abu EA, Ado SA, James DB. (2005). Raw starch degrading amylase production of mixed culture of *Aspergillus niger* and *Saccharomyces cerevisiae* grown on Sorghum pomace, *Afr. J. Biotechnol.* 4; 785-790.
4. Damai, S. (1994). Comparative study of *Aspergillus* species occurring on different plant seeds. Ph.D. Thesis, Dr. Babasaheb Ambedkar Marathwada Uni. Aurangabad.
5. Damien M, Catherine J, Patrice D, Christopher B (2010). Enhanced mechanical properties

- of partially beta-amylase trimmed starch for material application. *Carbohydr. Poly.* 80(3): 747-752.
6. Gupta, R., Giras, P., Mohapatra, H., Gaswami, Y.K., Chauhan, B. 2009. Microbial  $\alpha$ -amylase: a biotechnological perspective. *Process Biochem.*, 38(11): 1599-1616.
  7. Ikenebomeh M.J. and Chikundu A.E. (1997) *Aspergillus niger* Biomass production in cassava whey medium, *Nig.J. Microbiol.*, 11: 52- 63.
  8. Reeta Rani, Anil Kumar, Carlos R, Soccol Ashok Pandey (2009). Recent advances in solid state fermentation, *Biochem. Eng. J.* 44(1): 13-18
  9. Kulkarni, A. U., Chavan A. M., Kasare, U. T., & Pawar, S.M., (2012). Effect of nitrogen and phosphorous sources on amylase production in seed born fungi of maize. *Current Botany* Vol3, no.4.
  10. Hernandez M.S., Marilu R.R., Nelson P.G. and Renato P.R.(2006). Amylase production by *Aspergillus niger* in submerged cultivation on two wastes from food industries. *J. Food Eng.* 73: 93-100.
  11. Sivaramakrishnan, S., D. Gangadharan, K.M. Nampoothiri, C.R. Soccol and A. Pandey. (2006). Alpha amylases from microbial sources. An overview on recent developments. *Food Technol. Biotechnol.*, 44:173-184.
  12. S.L. Wang, Y.C. Liang, and T.W. Liang (2011). Purification and characterization of a novel alkali-stable  $\alpha$ -amylase from *Chryseobacterium taeanense* TKU001, and application in antioxidant and pre biotic, *Journal of Process Biochemistry*, vol. 46, no. 3, pp. 745-750.
  13. Sulochana R. Rathod and Ashok M.Chavan (2010). Extracellular Hydrolytic Enzymes Action of *Alternaria* Species under the Influence of Different Nutritional Sources. *Journal of Ecobiotechnology* 2/6: 57-62.
  14. Van der Maarel, M.J.E.C., B. Van der Veen, J.C.M. Vitdehaag, H. Leemhuis and L. Dijkhuizen. (2002). Properties and application of starch converting enzymes of the  $\alpha$ -amylase family. *Journal of Biotechnology*, vol 49: 137- 55.